

Emerging and endemic arboviruses: Challenges for laboratory surveillance

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Emerging diseases

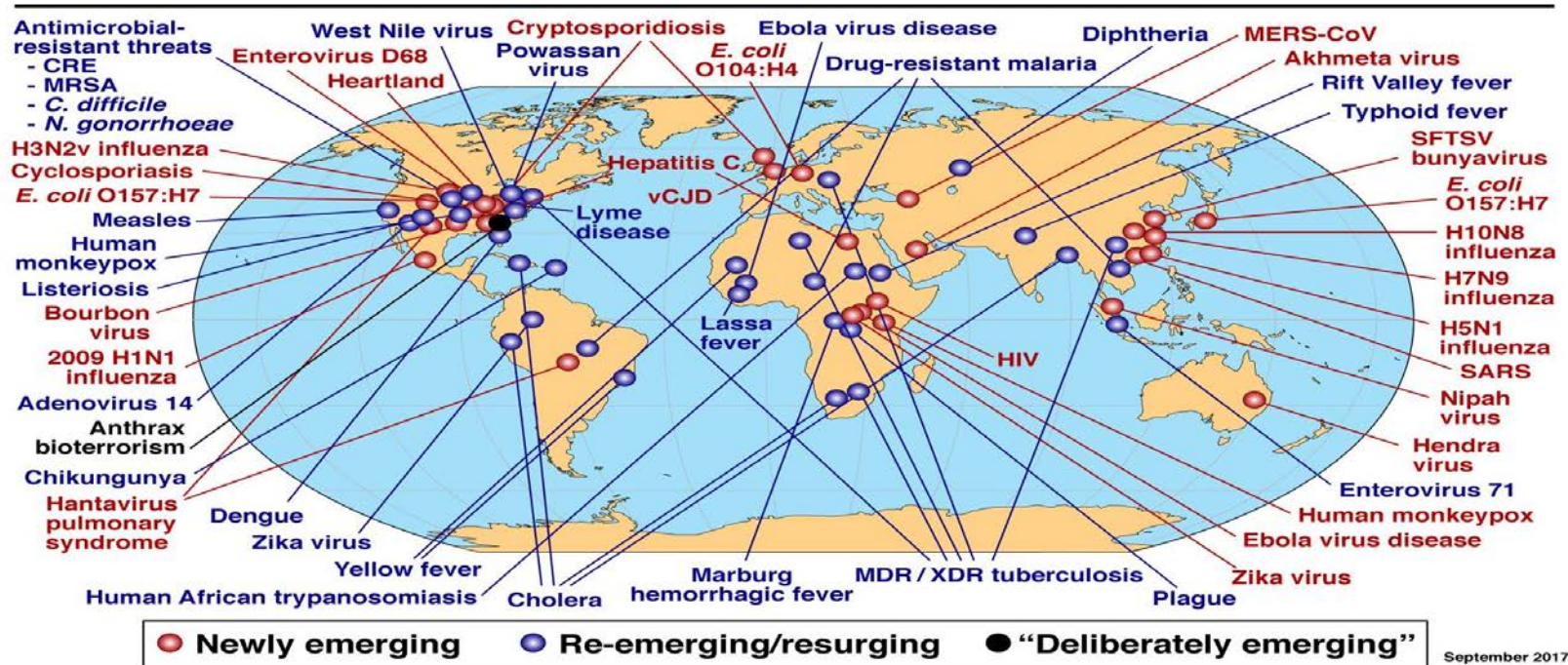
Microbial Threats to Health in the United States

1992, Instituto de Medicina (USA)
(Joshua Lederberg & Robert Shope)

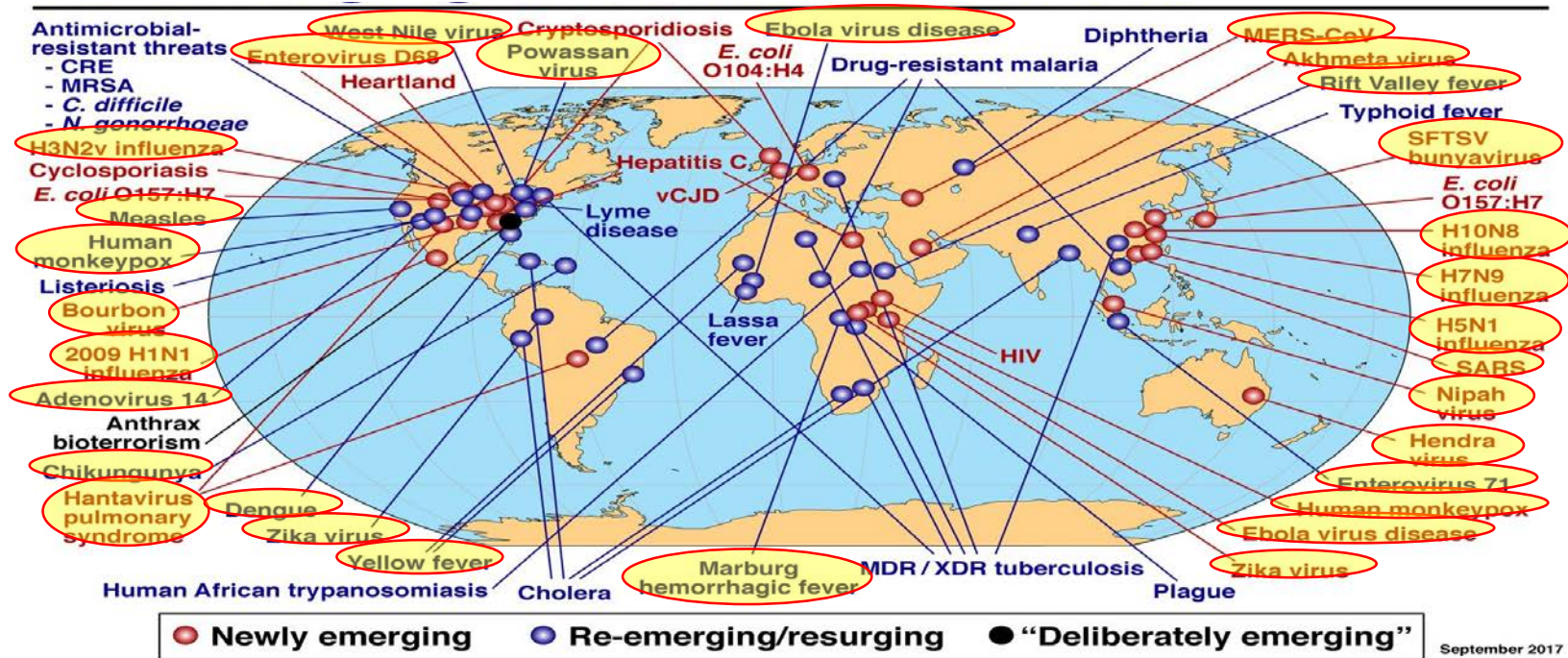
- ***Emerging diseases:***
new diseases / new pathogens (or new strains) / expansion to new territories
- ***Re-emerging diseases:***
diseases previously “under control” but re-appear increasing the incidence



Emerging & re-emerging diseases, 1996-2017



Emerging & re-emerging diseases, 1996-2017



Emerging Arboviruses



Epidemiological Alert

Zika virus infection
7 May 2015

The Pan American Health Organization (PAHO) / World Health Organization (WHO) recommends its Member States establish and maintain the capacity for Zika virus infection detection, clinical management and an effective public communication strategy to reduce the presence of the mosquito that transmits this disease, particularly in areas where the vector is present.



Epidemiological Alert

Neurological syndrome, congenital malformations, and Zika virus infection. Implications for public health in the Americas
1 December 2015

Given the increase of congenital anomalies, Guillain-Barré syndrome, and other neurological and autoimmune syndromes in areas where Zika virus is circulating and their possible relation to the virus, the Pan American Health Organization / World Health Organization (PAHO/WHO) recommends its Member States establish and maintain the capacity to detect and confirm Zika virus cases, prepare healthcare facilities for the possible increase in demand at all healthcare levels and specialized care for neurological syndromes, and to strengthen antenatal care. In addition, Member States should continue efforts to reduce the presence of mosquito vectors through an effective vector control strategy and public communication.



Epidemiological Alert

Increase of microcephaly in the northeast of Brazil

17 November 2015

Given the unusual increase in cases of microcephaly in some northeast states of Brazil, the Pan American Health Organization (PAHO) / World Health Organization (WHO) calls upon Member States to remain alert to the occurrence of similar events in their territories and to notify its occurrence through the channels established under the International Health Regulations (IHR).

Media centre

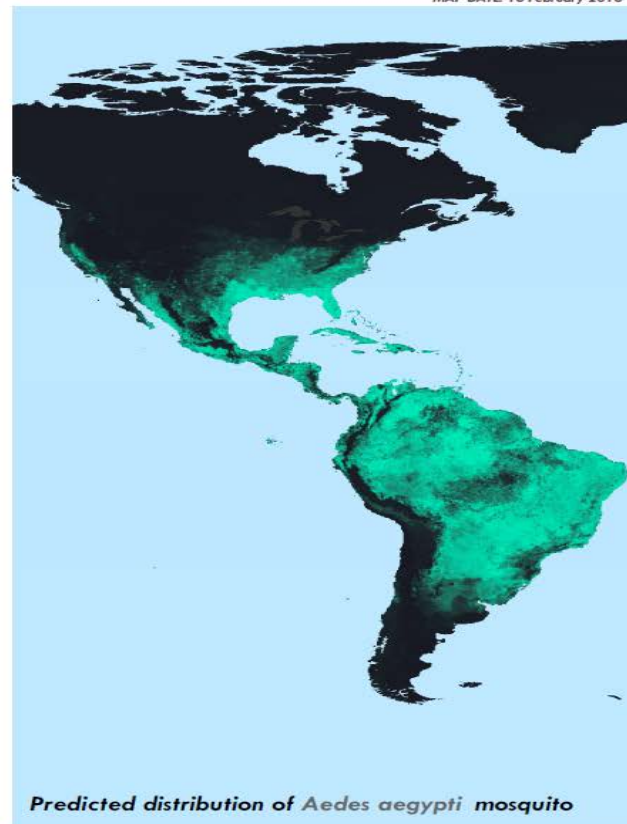
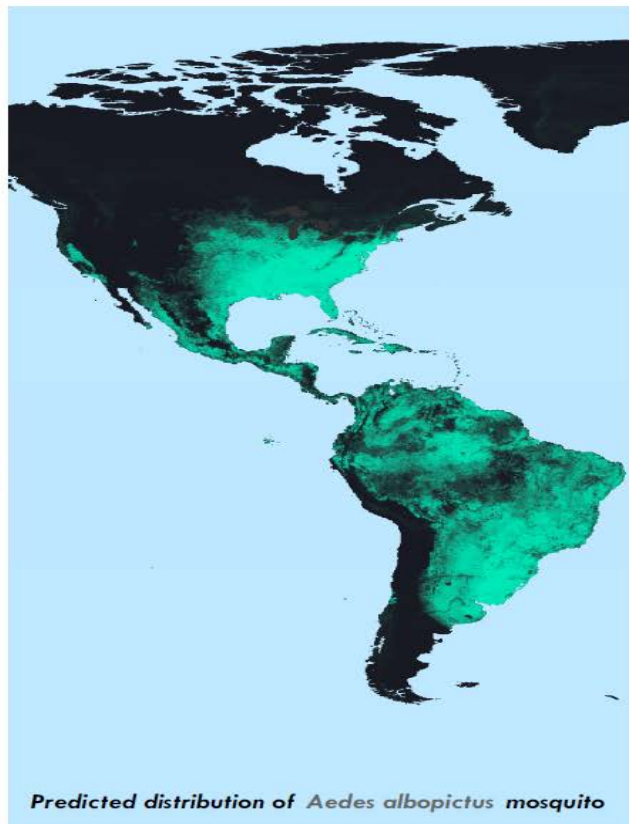
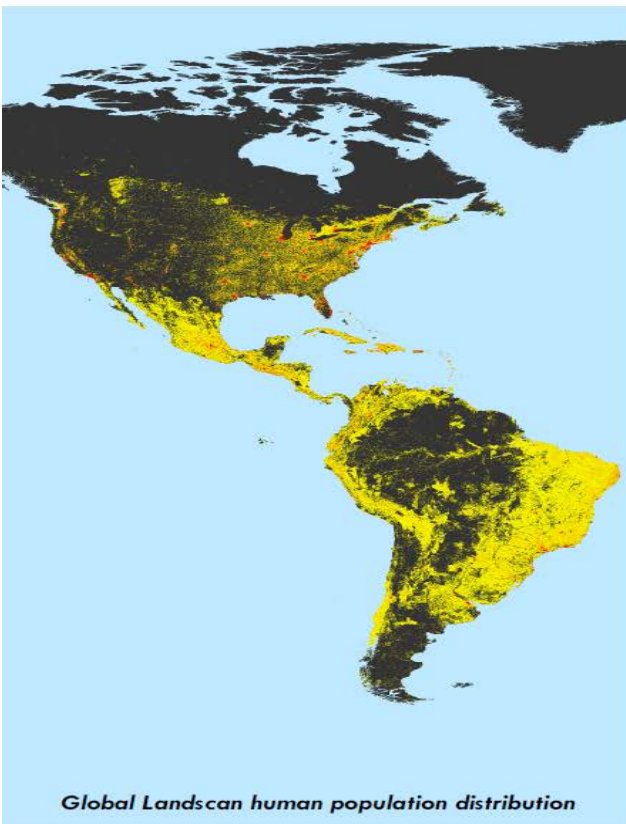
WHO statement on the first meeting of the International Health Regulations (2005) (IHR 2005) Emergency Committee on Zika virus and observed increase in neurological disorders and neonatal malformations

WHO statement
1 February 2016

Based on this advice the Director-General declared a Public Health Emergency of International Concern (PHEIC) on 1 February 2016. The Director-General endorsed the Committee's advice and issued them as Temporary Recommendations under IHR (2005). The Director-General thanked the Committee Members and Advisors for their advice.

Population density map and Predicted distribution of the *Aedes albopictus* and *Aedes aegypti* mosquito

MAP DATE: 16 February 2016



10 POPULATION PER SQ.KM. 5,000+



Population layer : Landscan 2010

High

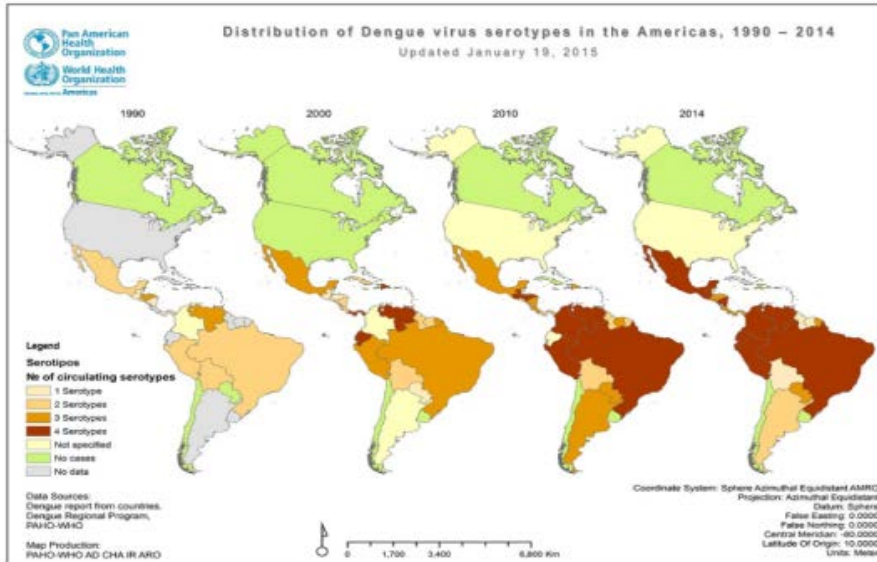


Low

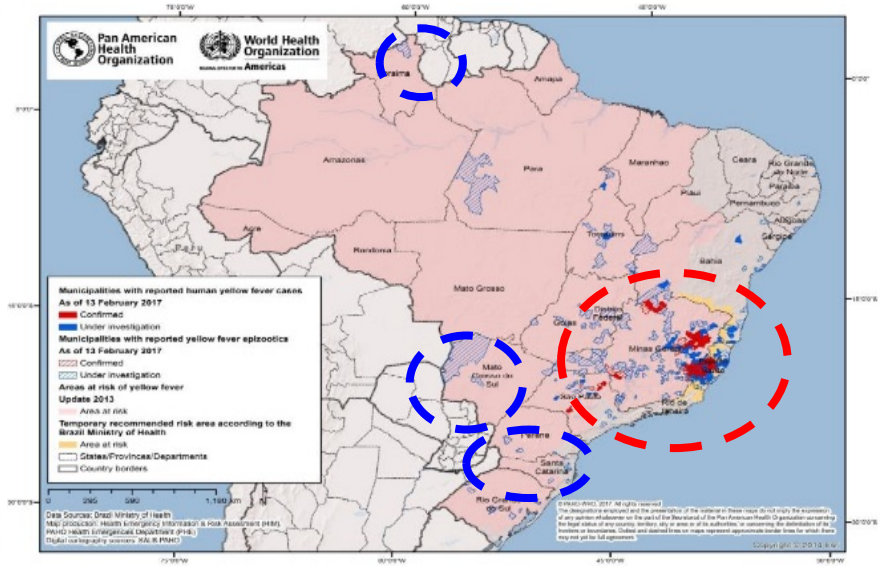
Service Layer Credits: Kraemer MUG et al. eLife

The boundaries and names shown and the designations used on this map do not imply the expression of any opinion whatsoever on the part of the World Health Organization concerning the legal status of any country, territory, city or area or of its boundaries, or concerning the distribution of its boundaries or territories. Dotted and dashed lines on maps represent approximate boundaries lines for which there may not yet be full agreement.

(Re) Emerging

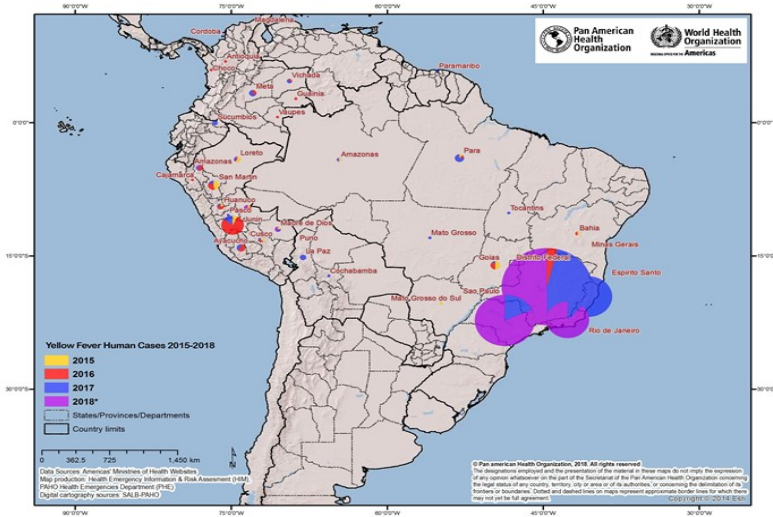


DENV
(endemic)



YFV (endemic,
re-emerging)

Yellow fever



2015: 27 cases in **Peru**, Brazil
 2016: 113 cases in **Peru**, Brazil, Colombia,
 2017: 819 cases in **Brazil**, Peru, Bolivia, Ecuador, Colombia, Suriname, Fr. Guiana

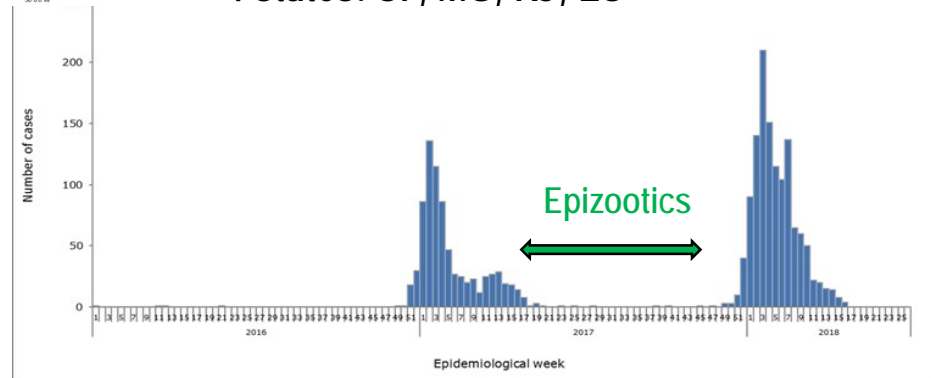
South-east Brazil:

December 2016 – June 2017

- 792 confirmed cases
- 274 deaths [CFR: 34.6%]
- 4 states: SP, MG, RJ, ES

*July 2017- June 2018**

- 1266 confirmed cases
- 415 deaths [CFR: 32.8%]
- 4 states: SP, MG, RJ, ES



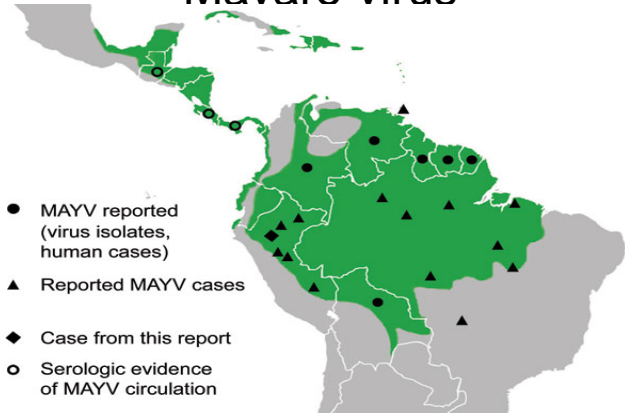
Distribution of confirmed yellow fever cases by epidemiological week (EW). Brazil, 2016 – 2018*

Other arboviruses affecting humans

Yellow Fever	<i>Flaviviridae</i>	Mosquitoes: <i>Aedes</i> and <i>Haemogogus</i> spp.	Primates	Europe, Oceania South America, Africa
West Nile	<i>Flaviviridae</i>	Mosquitoes: <i>Culex</i> spp	Birds, Horses, Other Mammals	Africa, Asia, Europe, Oceania, Americas
Japanese encephalitis	<i>Flaviviridae</i>	Mosquitoes: <i>Culex</i> spp	Birds, Pigs	Asia, Oceania
Murray Valley encephalitis	<i>Flaviviridae</i>	Mosquitoes: <i>Culex</i> spp	Birds	Oceania
Zika virus	<i>Flaviviridae</i>	Mosquitoes: <i>Aedes</i> spp	Primates	Africa, Asia, Oceania, Central and South America
Rocio	<i>Flaviviridae</i>	Mosquitoes: <i>Psorophora</i> and <i>Aedes</i> spp	Birds	South America
St. Louis encephalitis	<i>Flaviviridae</i>	Mosquitoes: <i>Culex</i> spp	Birds, Bats, Other Mammals	Americas
Kyasanur Forest disease	<i>Flaviviridae</i>	Ticks: <i>Hemaphysalis</i> spp.	Primates, Rodents, Other Mammals	Asia
Omsk hemorrhagic fever	<i>Flaviviridae</i>	Ticks: <i>Dermacentor</i> and <i>Ixodes</i> spp Mosquitoes: ?	Rodents, Voles, Other Mammals	Europe
Tick-borne encephalitis	<i>Flaviviridae</i>	Ticks: <i>Ixodes</i> spp	Rodents, Goats, Sheep, Cows, Other Mammals, Birds?	Europe, Asia
Sandfly fever	<i>Bunyaviridae</i>	Sandflies: <i>Phlebotomus</i> spp.	Birds? Mammals?	Europe, Asia, Africa
Rift Valley fever	<i>Bunyaviridae</i>	Mosquitoes: <i>Aedes</i> , <i>Ochlerotatus</i> , <i>Stegomyia</i> , <i>Anopheles</i> , <i>Culex</i> , <i>Neomelanimonion</i> , <i>Eretmapodites</i> and others	Cows, Sheep, Camels, Goats and Other Mammals	Africa, Asia
La Crosse encephalitis	<i>Bunyaviridae</i>	Mosquitoes: <i>Aedes</i> spp	Rodents	North America
Crimean-Congo hemorrhagic fever	<i>Bunyaviridae</i>	Ticks: <i>Hyalomma</i> spp	Cows, Sheep, Goats, Hares and Other Mammals	Europe, Asia, Africa
Oropouche	<i>Bunyaviridae</i>	Midges: <i>Culicoides</i> sp	Primates? Sloths? Birds?	Central and South America
Severe febrile thrombocytopenia syndrome	<i>Bunyaviridae</i>	Ticks: <i>Haemaphysalis</i> sp	?	Asia
Chandipura	<i>Rhabdoviridae</i>	Sandflies: <i>Phlebotomus</i> and <i>Sergentomyia</i> spp.	Hedgehogs, Others?	Asia and Africa
Bluetongue	<i>Reoviridae</i>	Midges: <i>Culicoides</i> spp	Sheep, Cows, Other Mammals	Africa, Asia, Europe, Oceania, Americas (all except Antarctica)

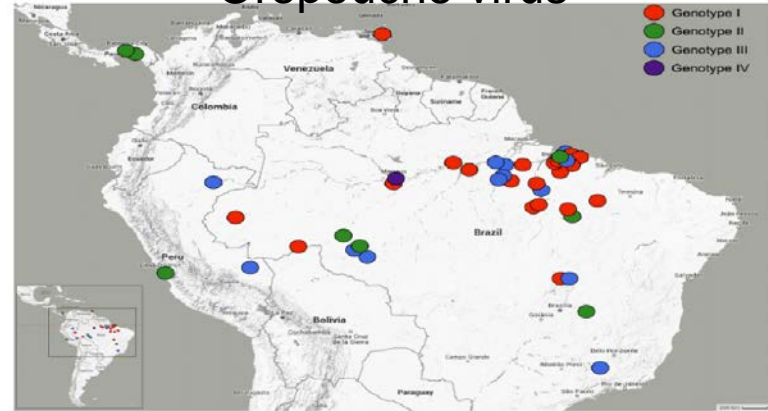
Mayaro and other arboviruses...

Mayaro virus



Emerg Infect Dis, 2012, 18:695-6

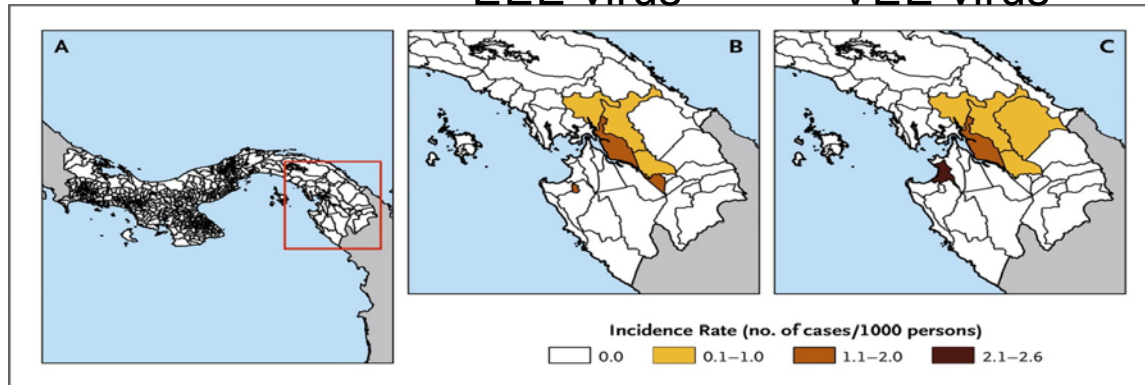
Oropouche virus



Am J Trop Med Hyg, 2017, 96:1019-30

EEE virus

VEE virus



N Engl J Med, 2013, 369:732-744

Surveillance and early response: Role of the laboratory

Surveillance and early response

International Health Regulations:

- Recommends countries to maintain active surveillance of diseases and public health events.
- Urges to strengthen and respond quickly to events of international dispersion and contain any threat to public health.

Pathogens under
elimination or control:
viruela, poliovirus, etc...

Emerging or re-emerging
pathogens (Zika, YFV,
SARS, MERS, Flu...)

Early Warning System



Surveillance and early response

International Health Regulations:

- Core capacity # 8 of the IHR (2005) **obligates** WHO Member States to establish mechanisms to provide **reliable and timely identification** and characterization of infectious agents and other hazards that may cause public health emergencies of national and international interest, including sending specimens to the appropriate laboratories if necessary

Pathogens under
elimination or control:
viruela, poliovirus, etc...

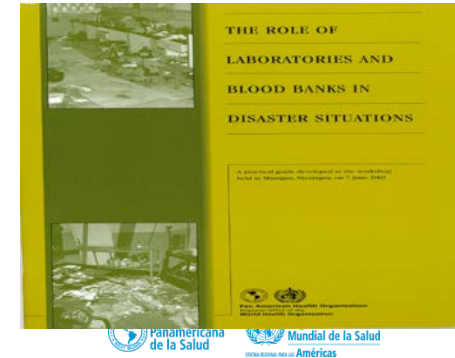
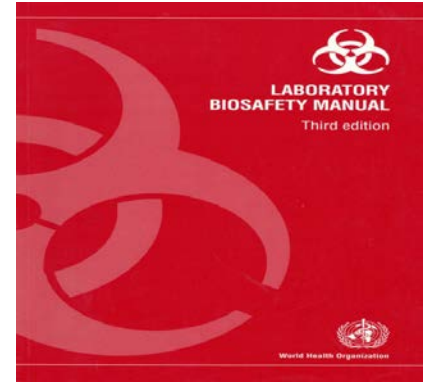
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Early Warning System



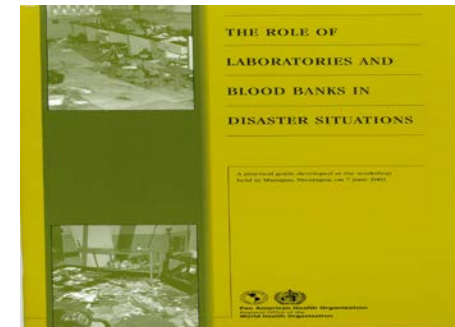
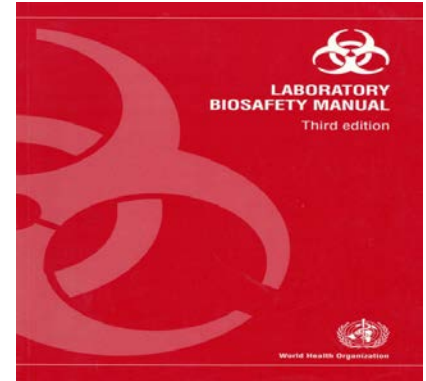
IHR, Core capacity # 8 : Critical laboratory elements

1. Capacity to diagnose priority pathogens
2. Quality management
3. Management of biological risk
4. Collection and transport of samples
5. Laboratory-based surveillance
6. Laboratory networks



IHR, Core capacity # 8 : Critical laboratory elements

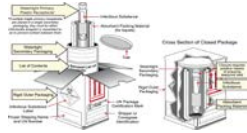
1. **Capacity to diagnose priority pathogens**
2. Quality management
3. Management of biological risk
4. Collection and transport of samples
5. Laboratory-based surveillance
6. Laboratory networks



Standardization of processes

Processing

- Type of sample
- Sample conservation
- Sample transportation
- Type of test
- Equipment and Reagents
- Reference materials

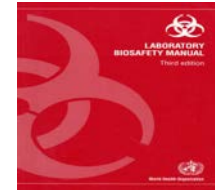


Algorithms

- Sequential Vs simultaneous
- Singleplex Vs Multiplex
- Differentials

Implementation

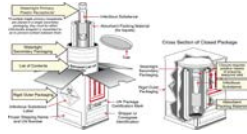
- Training
- SoPs
- Equipment calibration
- Interpretation of results
- Limitations
- QC
- EQAP
- Biosecurity
- Biosafety
- Maintenance
- Waste disposal



Standardization of processes

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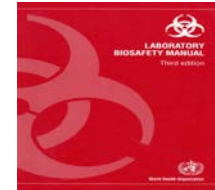


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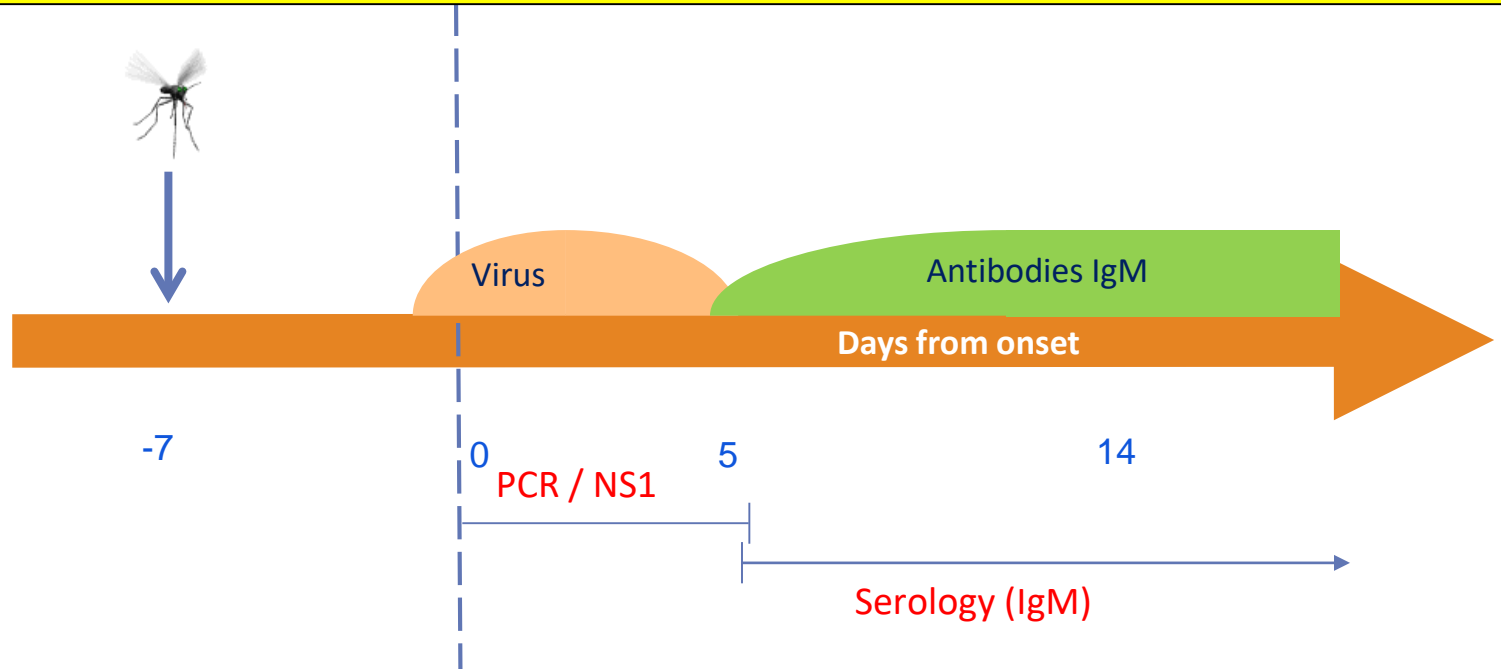
- Training
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Interpretation

Laboratory algorithms

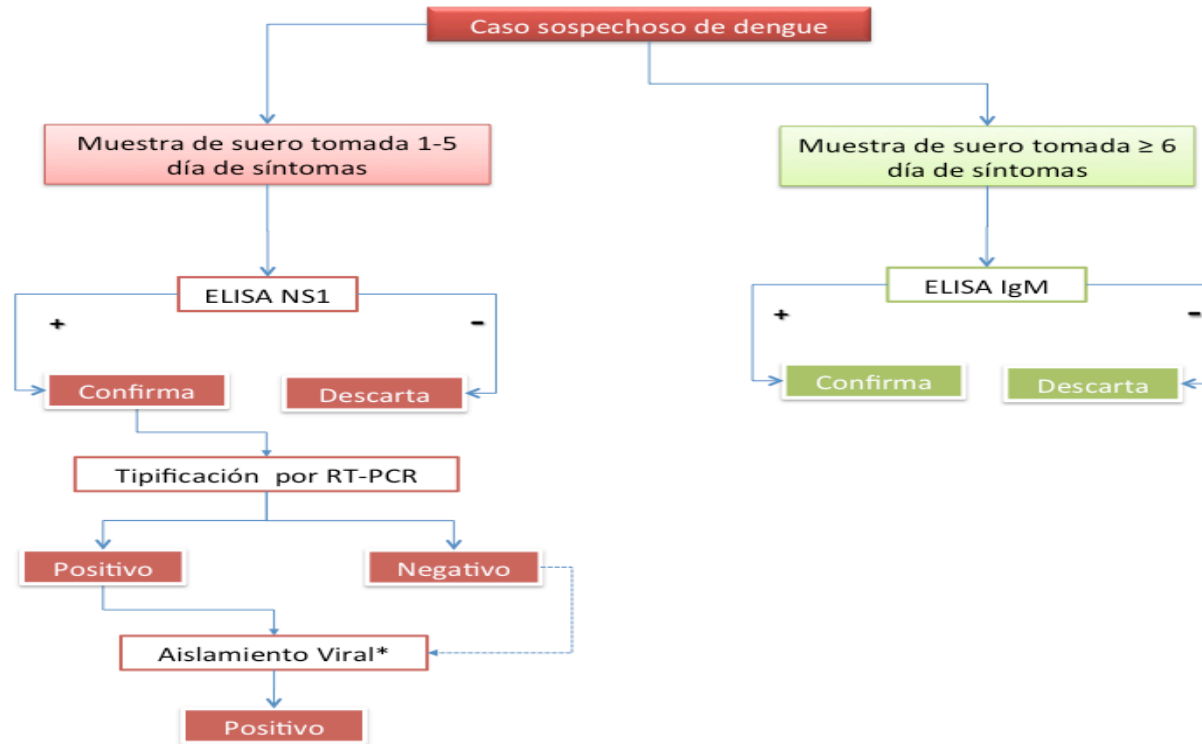
Infection dynamics: Replication / Immune response



Processing algorithms

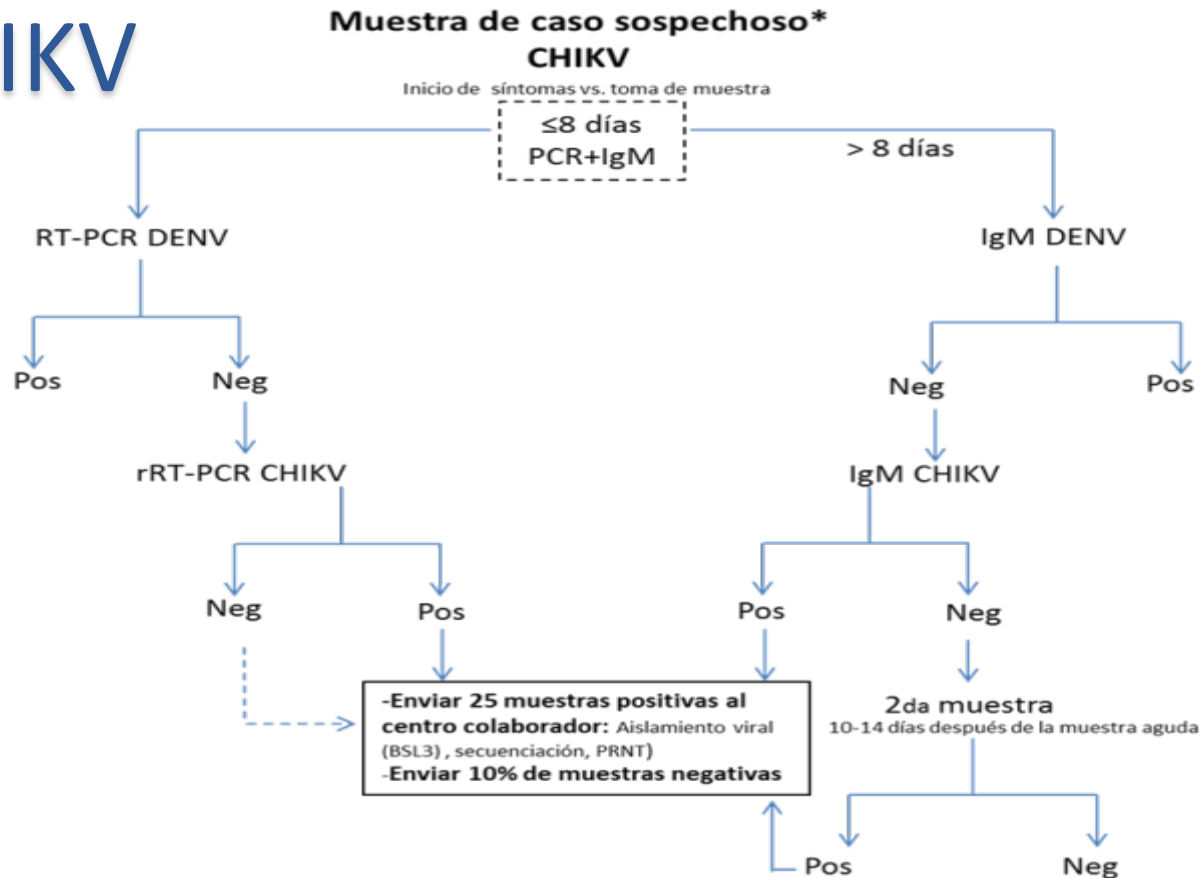
- The laboratory algorithms are **NOT** static and should be adjusted depending on the needs, epidemiological profile and to respond to emergencies

Dengue

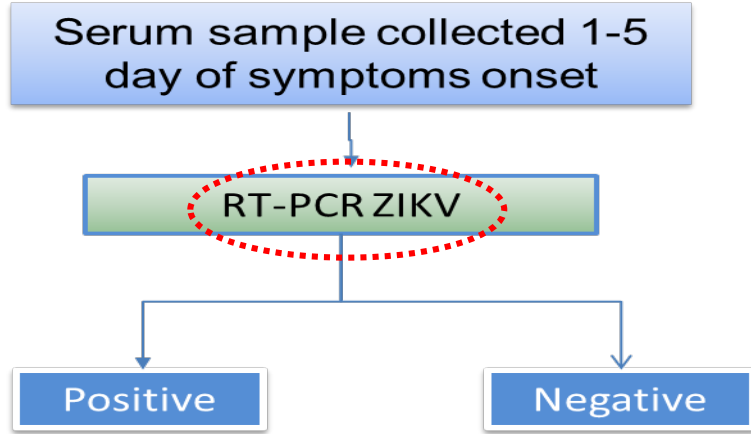


Adaptado de: Laboratorio de Arbovirus-Grupo Virología. INS Colombia

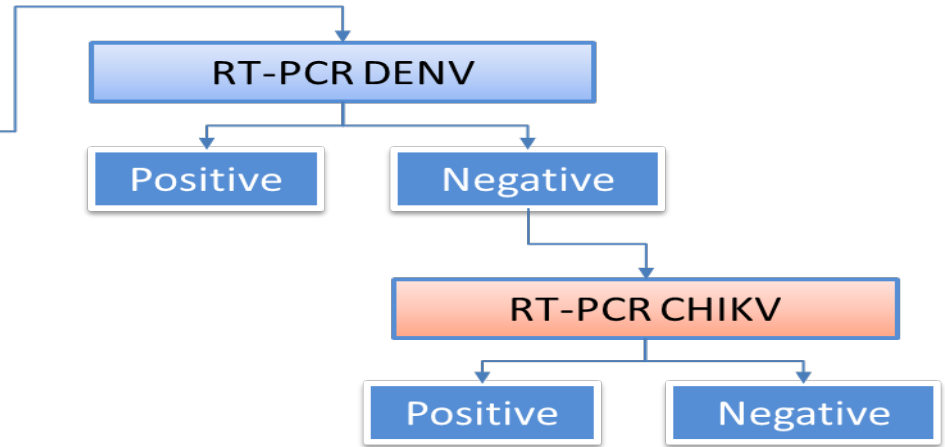
CHIKV



Algorithm for *integrated* surveillance



Algorithm A: Sequential



Algorithm for *integrated* surveillance

Algorithm B: Parallel

Serum sample collected 1-5 day of symptoms onset

ZIKV

RT-PCR ZIKV

Positive

Negative

DENV

RT-PCR DENV

Negative

Positive

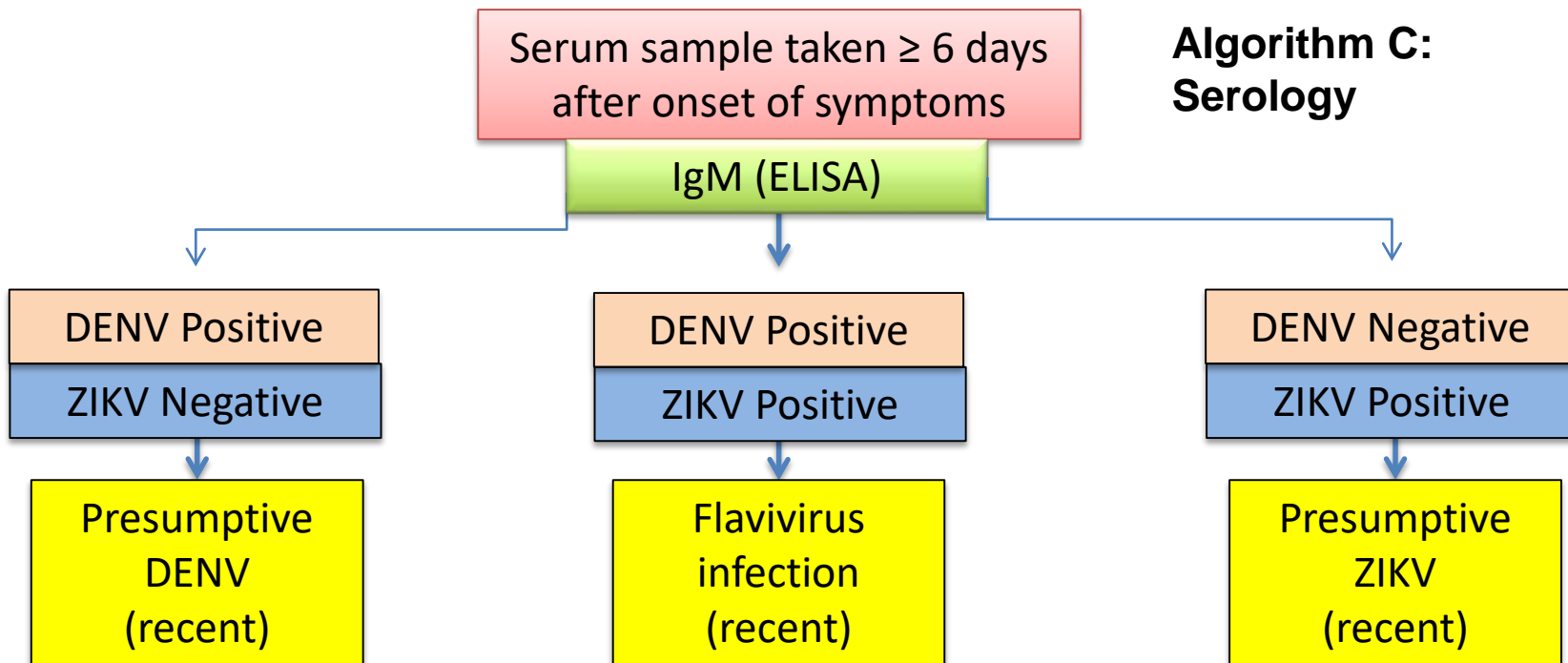
CHIKV

RT-PCR CHIKV

Negative

Positive

Algorithm for *integrated* (serological) surveillance



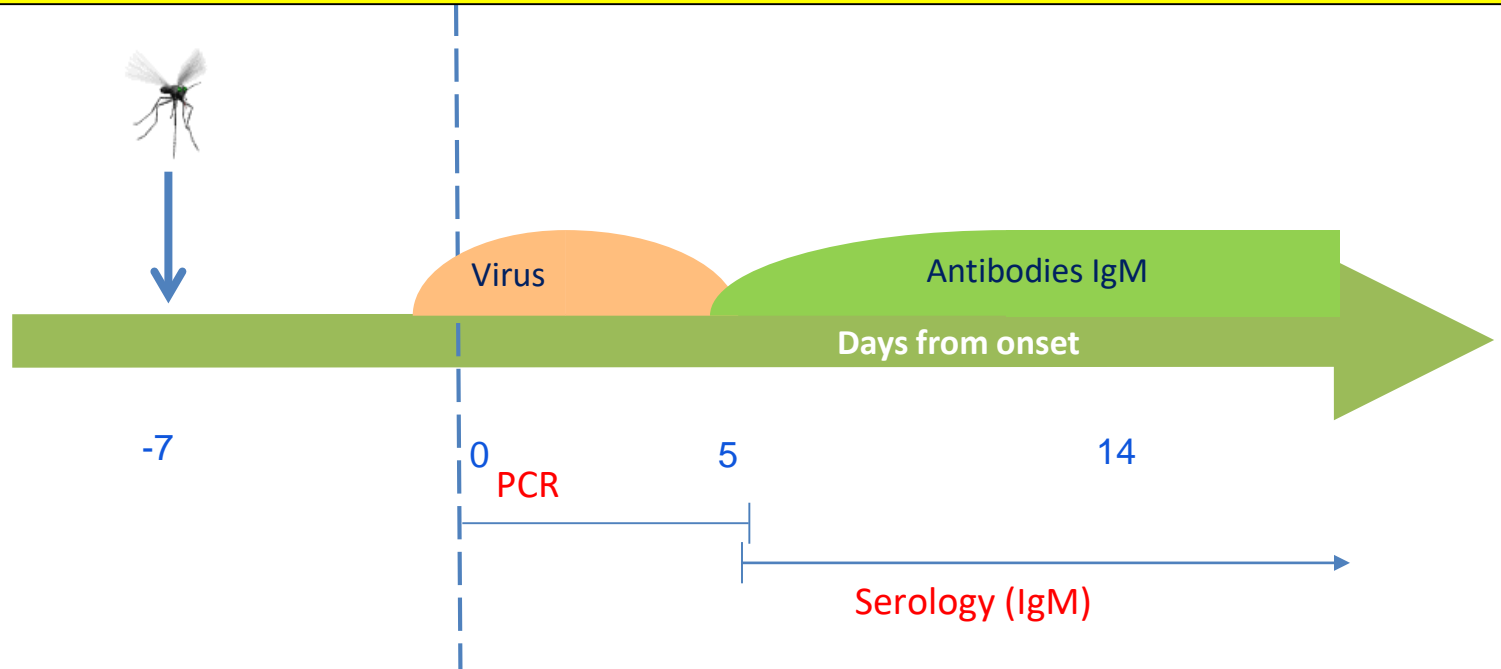
Processing algorithms

- The laboratory algorithms are **NOT** static and should be adjusted depending on the needs, epidemiological profile and to respond to emergencies

Take into account new findings, biological evidence and performance of new assays

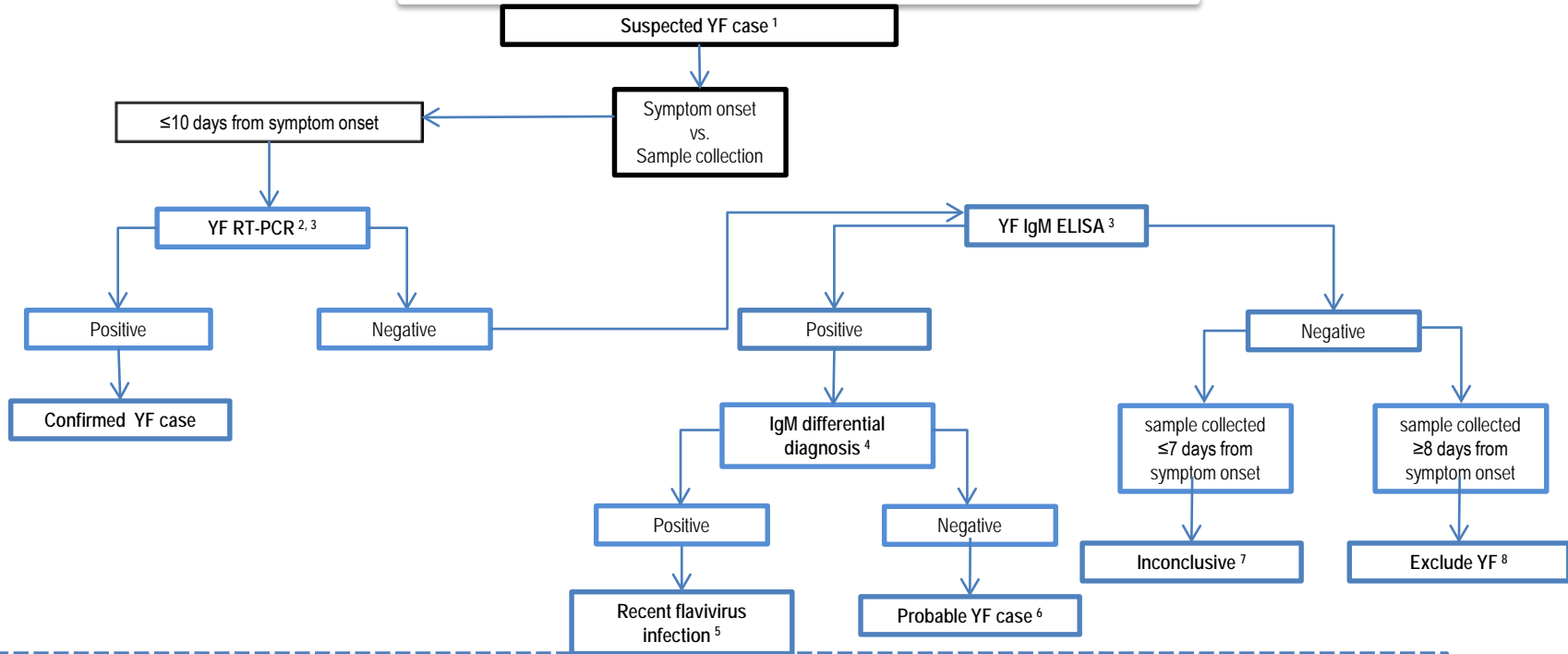
Laboratory algorithms

Infection dynamics: Replication / Immune response



Slide adapted from Dr. Jorge Muñoz, CDC Pto. Rico

Algorithm for laboratory confirmation of yellow fever (YF) cases



¹ No YF vaccination within 30 days or unknown YF vaccination history.

² Laboratories that only have the capacity to perform RT-PCR or IgM ELISA should test samples with the available technique. Results should be interpreted according to the algorithm.

³ RT-PCR sensitivity is higher in the first 10 days from symptom onset. However, detection beyond 10 days has been reported, in particular in severe (and fatal) cases.

⁴ Must include dengue virus as well as other flaviviruses depending on the epidemiological situation of the area/country.

⁵ Consider performing PRNT in a reference laboratory. This result does not rule out yellow fever. Thus, in areas where no YF circulation has been described recently, this should prompt an investigation.

⁶ A positive IgM test in a single sample is not confirmatory. Additional clinical and epidemiological criteria must be used for the final interpretation of the case, in particular in areas where no YF circulation has been described recently.

⁷ A second sample should be requested and tested according to the algorithm.

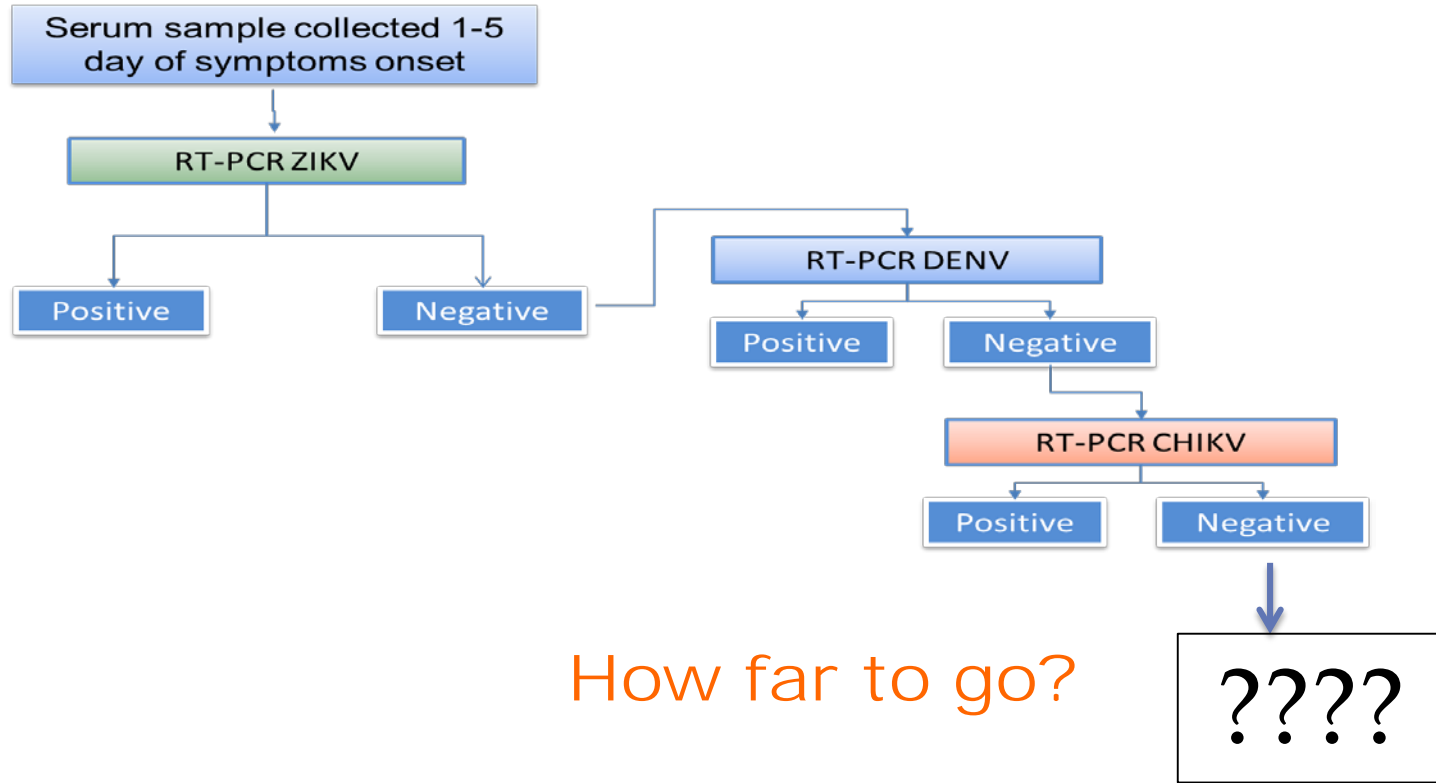
⁸ Cases should be investigated and clinical differential diagnosis performed.

Processing algorithms

- The laboratory algorithms are **NOT** static and should be adjusted depending on the needs, epidemiological profile and to respond to emergencies

For early detection of emerging agents,
the **negative** samples are as important as the positive ones

Algorithms to detect emerging arboviruses...



How far to go?



Organización
Panamericana
de la Salud



Organización
Mundial de la Salud
OFICINA REGIONAL PARA LAS Américas

Algorithms to detect emerging arboviruses...

- Additional assays...(*singleplex?*)
- Generic assays
 - *Panflavivirus*
 - *Panalphavirus*
 - *Panbunyavirus*

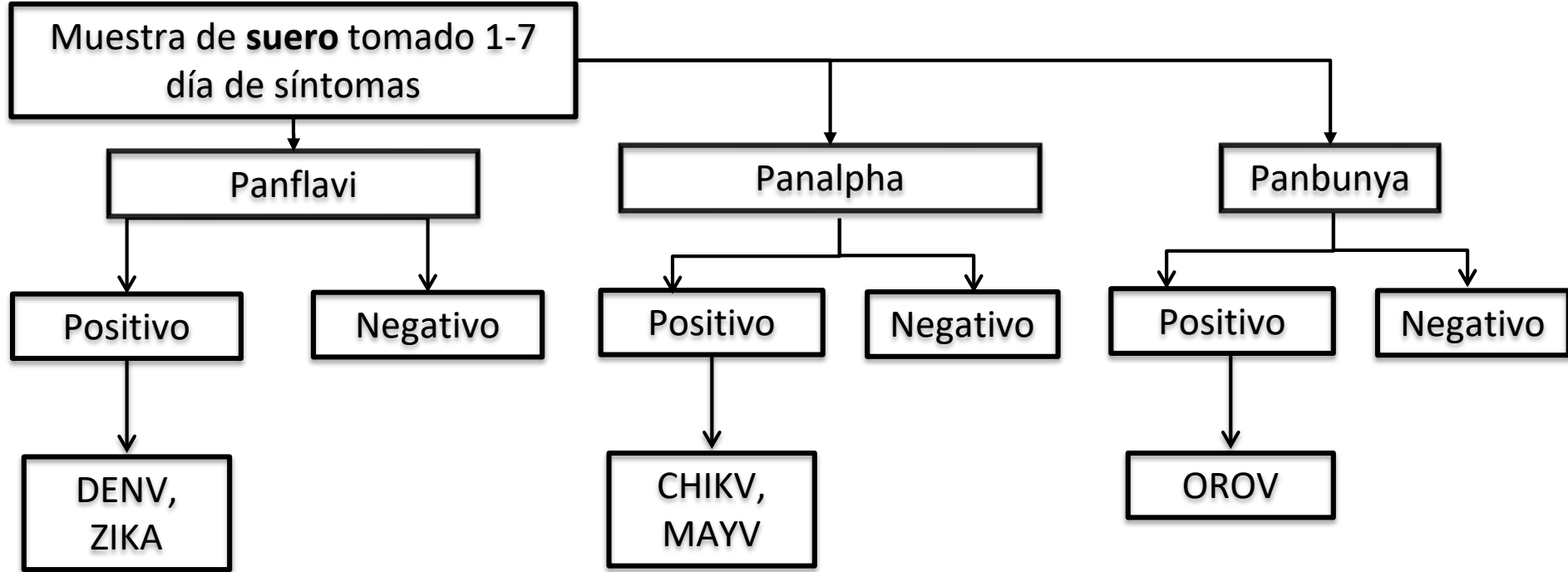


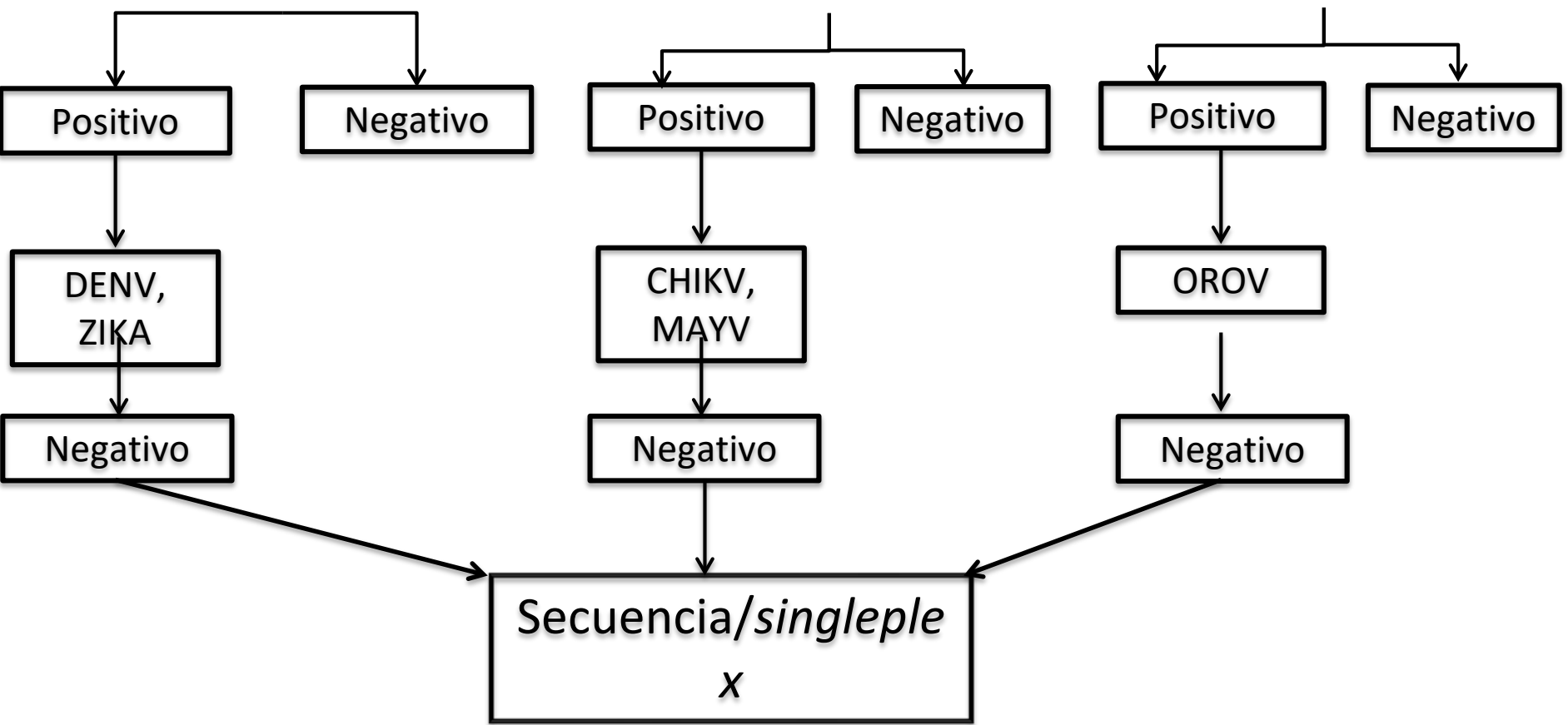
Pan American
Health
Organization



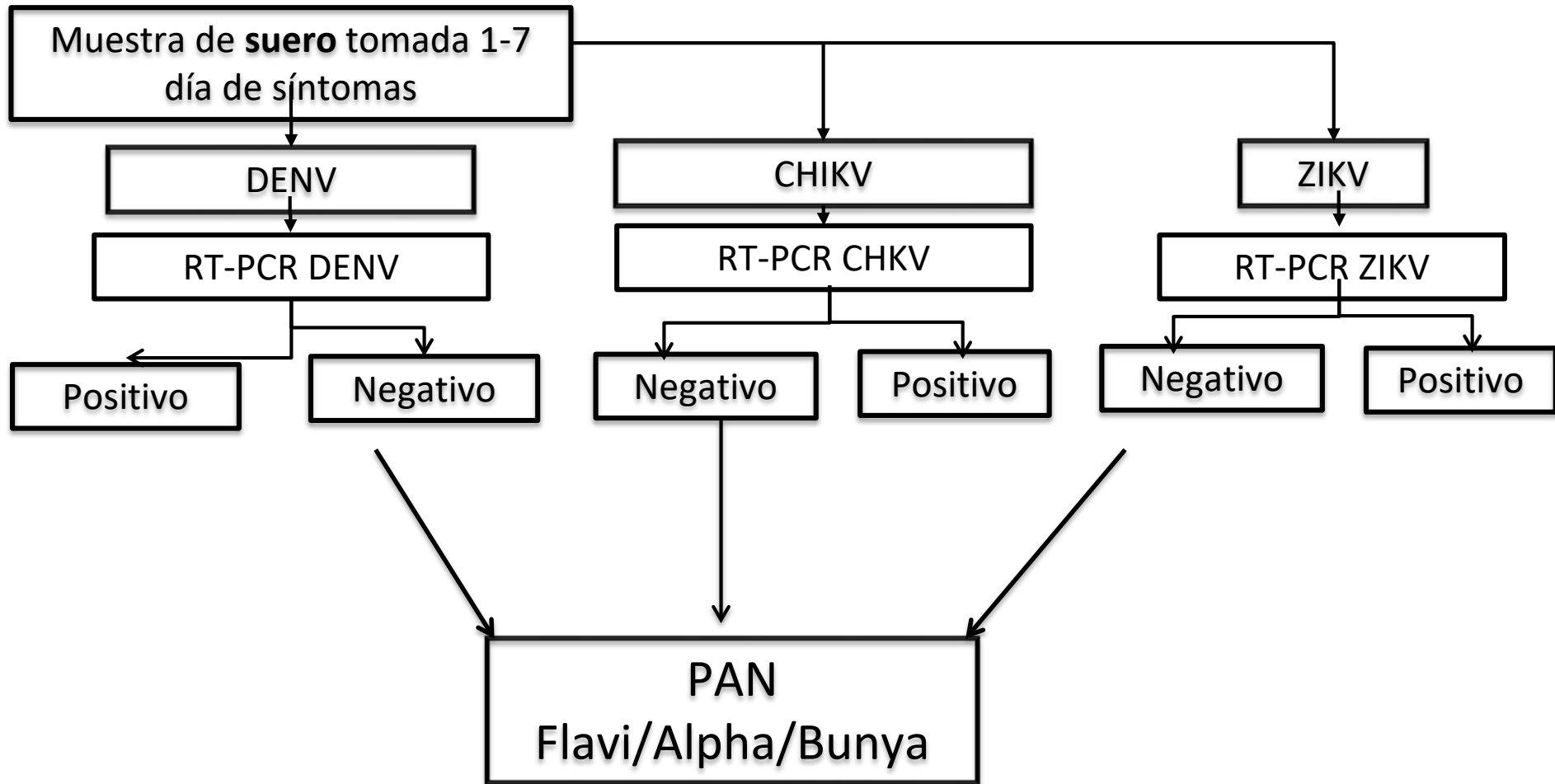
World Health
Organization
REGIONAL OFFICE FOR THE
Americas

“Generic” algorithms (1)

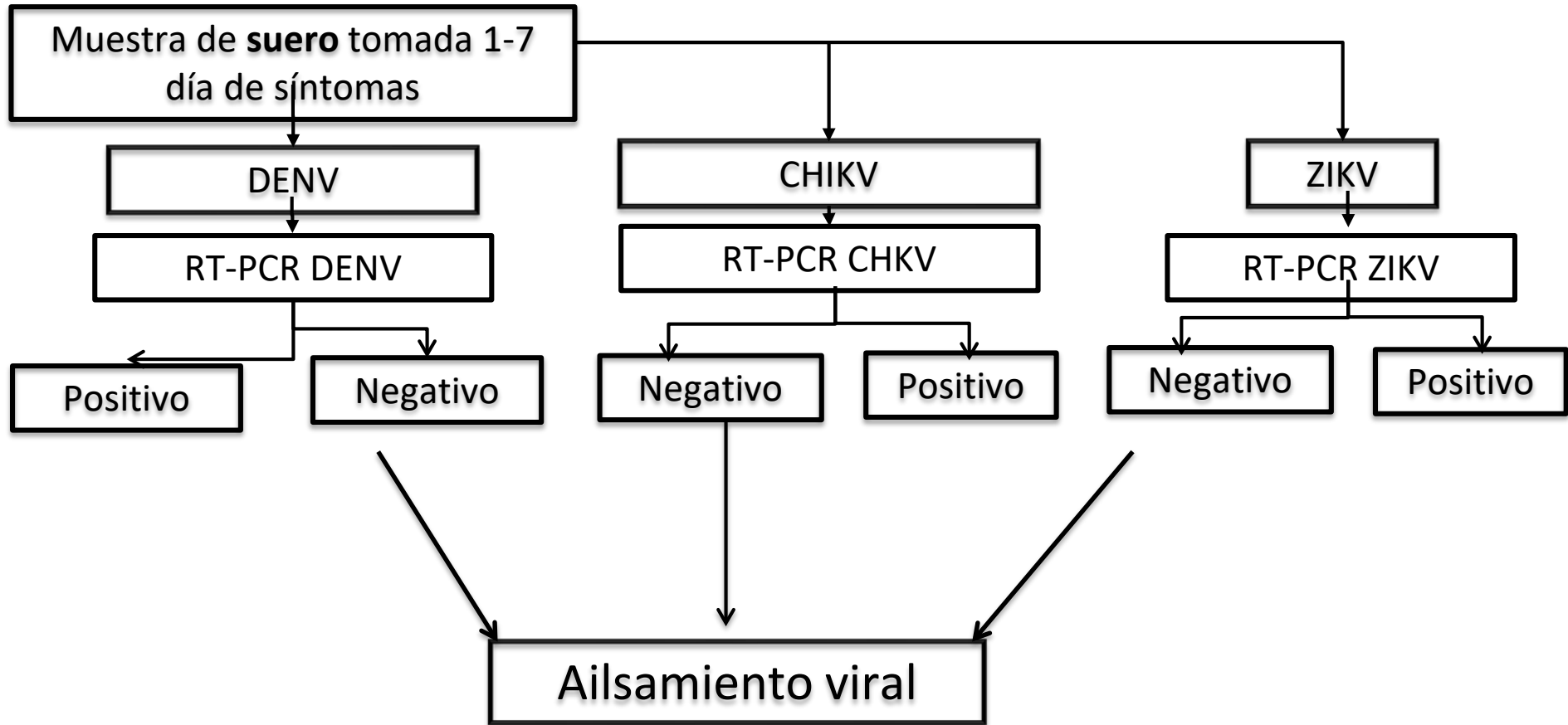




“Generic” algorithms (2)



“Generic” algorithms (3)

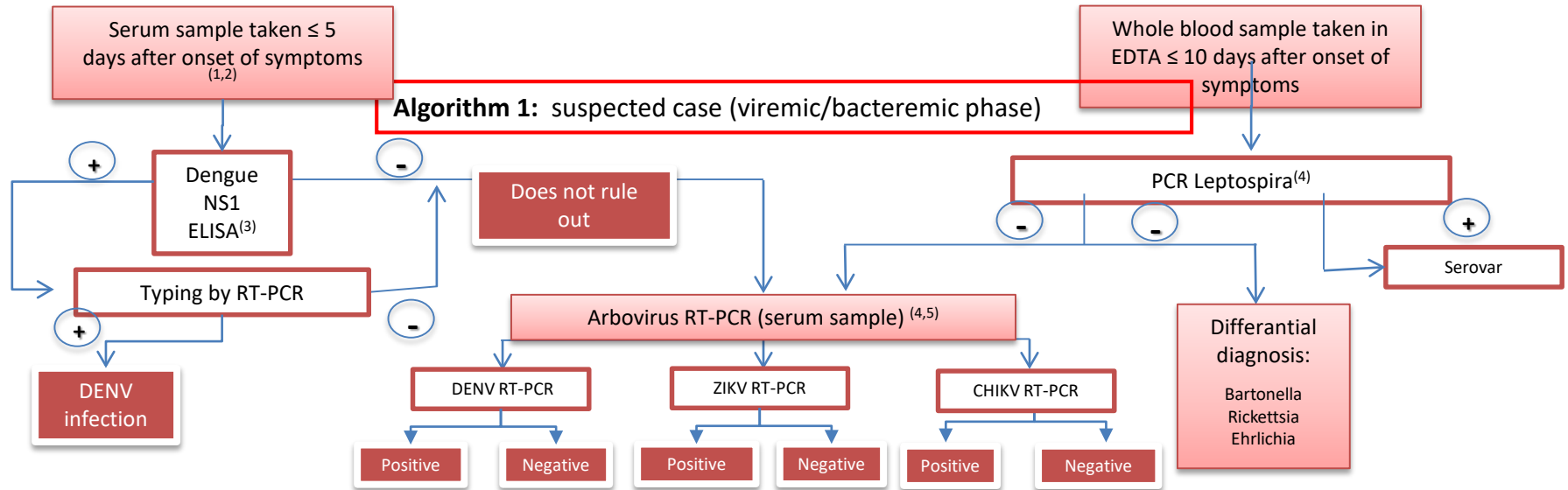


Processing algorithms

- The laboratory algorithms are **NOT** static and should be adjusted depending on the needs, epidemiological profile and to respond to emergencies

In case of emergencies due to natural disasters, the differential diagnosis must be considered with other types of agents

Algorithm for the differential diagnosis of arbovirus and leptospira infection in areas of documented co-circulation and post-emergency areas at risk



- (1) Perform a leptosira IgM ELISA with this serum sample and a second sample collected at least 14 days after the first to test for seroconversion
- (2) A minimum sample volume of 5ml should be collected to obtain at least 1ml of serum for arboviruses testing and 1ml of serum for leptospira testing
- (3) Sensitivity can vary depending on the serotype
- (4) Molecular detection can be performed sequentially (*singleplex*, starting with the most probable agent according to clinical criteria) or in parallel (*multiplex*)
- (5) ZIKV can also be detected by RT-PCR in urine from day 1 to day 15 (on average)

Processing algorithms

- The laboratory algorithms are **NOT** static and should be adjusted depending on the needs, epidemiological profile and to respond to emergencies

For the differential diagnosis of other pathologies considered in the IHR and that are in the process of elimination: Measles vs Zika

Proposed algorithm for Zika surveillance integrated to Measles

Paciente con fiebre y rash detectado por el sistema de vigilancia S/R (según definición de caso)

Colectar muestra de orina y suero

Procesar S/R

+

Continuar estudio S/R

-

Algoritmo detección ZIKV

Muestra de suero ≤ 5 días
Algoritmo A or B

Muestra de suero ≥ 6 días
Algorithm C

Comentarios finales

- Adequate surveillance allows monitoring endemic pathogens
- Efficient mechanisms are required to identify new events, new agents (viruses), or new variants with pandemic potential.

SURVEILLANCE OF UNUSUAL CASES

Comentarios finales

- The laboratory is critical to confirm (or rule out) new agents: **Zika, Mayaro, Oropouche, EEV, West Nile ...**
- LSPs must be prepared to detect and report new agents in a timely manner (mandatory notification within 24 hours, RSI)
- However, the detection capacity does not refer only to the installed capacity; It implies the possibility of having access to a laboratory that has the capacity (**Networking!**)

Comentarios finales

- A good laboratory diagnosis depends on a **good sample** and a well recognized case ...
- Surveillance results **should not be used / expected to make medical decisions!** (the clinical diagnosis should be prioritized!)
- Articulation of the laboratory with the epidemiology and clinical components is essential to ensure an appropriate response to the IHR

Thank you!

Jairo A. Méndez-Rico, PhD

Regional Advisor Viral Diseases

PAHO/PHE